Yeast and Mold	Orange Serum Media	Wallerstein Nutrient Broth (WLN)	m-Green/Schauffus Pottinger Media	Yeast & Mold Broth	Brettanomyces Broth	PRY (Preservative Resistant Yeast) Broth	Wort Media	Morris Orange Serum	Yeast and Mold Selective Broth		
Catalogue Number	MHA000P20 (50/Pk)	MHA00P2N (50/Pk)	MHA000P2M (50/Pk)	MX00YM220 (20/Pk)	MHA00BSM2 (50/Pk)	MHAOOPRY2 (50/Pk)	NPWYM0150 (150/Pk)	MHA00MS02	MHA00P2SM (with Chloramphenicol) (50/Pk)		
Application	Used for the isolation and enumeration of acidophilic and aciduric microorganisms in water, beverages and foods.	Used to detect yeast and mold in worts, beers and other fermentation products. Used in cojunction with WLD.	A low pH culture medium used to detect yeast and mold and other aciduric organisms	Used for the isolation and cultivation of yeast, mold and other aciduric organisms.	Detection of <i>Brettanomyces</i> in wine and beer. Bacteria and other yeasts are inhibited.	A low pH selective culture medium for the detection of spoilage microorganisms resistant to acetic acid.	Used for the isolation and enumeration of yeast in beverages, beer and wine.	Used to detect yeast and mold resistant to sodium benzoate (like <i>Zygosaccharomyces bailii</i>) in beverage.	Used to detect yeast and mold. Chloramphenicol is used to suppress background bacterial growth.		
Incubation Time & Temperature	48 hrs – 5 days at 24 – 32°C	48 hrs – 5 days for yeast at 20°C 35°C for bacteria	48 – 72 hrs at 28 – 32°C	48 – 72 hrs at 20 – 30°C	5 – 7 days at 25°C	48 - 72 hrs at 30°C	48 hrs - 5 days at 22.5°C	2 -7 days at 25-30°C	48 – 72 hrs at 28 – 35 °C		
Typical Colony Appearance	Yeast appear white, creamy and large. Bacteria are smaller, white or transparent.	Mold can appear non-pigmented to white, with various texture. Yeast appear as creamy, white larger colonies. Bacteria appear blue-green.	Yeast are large green opaque colonies. Mold appears green and filamentous. Bacteria able to grow at this pH form smaller clear to white colonies.	Yeast produce white colonies with a creamy texture. Mold is rough textured and/or filamentous. Bacteria are smaller and clear to white.	Colonies appear small, white and creamy.	Yellow	Yeasts develop smooth white or colored colonies.	Yeast colonies appear as white, creamy and large colonies.	Yeast appear as large green and opaque colonies. Mold is green and filamentous.		
pH at 25 °C	MHA000P20: 5.6 \pm 0.2 NPOSA0150: 4.5 \pm 0.2 (with time, a slight reduction of pH may be noticed, which will not affect the recovery performance of the product)	5.5 ±0.2	MHA000P2M: 4.6 ±0.2 NPSPY0150: 4.5 ±0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	4.6 ±0.3	3.5 ±0.2	3.6 ±0.2	4.5 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	3.5 ± 0.2	4.4 ±0.2		
Packaging Type	MHA000P20: Non-luer tip ampoule NPOSA0150: Dehydrated nutrient pad in 47mm dish	MHAOOP2N: (Non-luer tip ampoule) MXOOWN220: (Luer tip ampoule)	MHA000P2M: (Non-luer tip ampoule) NPSPY0150: (Dehydrated nutrient pad in 47mm dish)	Luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule	Non-luer tip ampoule		
Total Viable Organism/Total Viable Count	Heterotrophic Plate Count (HPC) Broth	Tryptone Glucose Extract (TGE) Broth	Tryptic Soybean Broth (TSB)	Standard Count Media							
Catalogue Number	MHA000P2S (50/Pk)	MHA000P2T (50/Pk)	MX00TT220 (20/Pk)	NPSTC0150 (150/Pk)			ТМ	Σ			
Application	Recovery of heterotrophic bacteria found in various	A non-selective medium to detect total	Used to detect total heterotrophic	With TTC Indicator: NPTTC0150 (150/Pk) Used for the cultivation of fastidious and other							
Incubation Time	types of water, especially high-purity or potable. Prior to use, warm the media at 30-50 °C until liquefied.	heterotrophic microorganisms in water and other liquids.	microorganisms in water and other liquids.	microorganisms found in water, wastewater, raw materials, beverages, beer, food, etc.							
& Temperature	40 72 IIIS at 25 35 C	40 - 721115 at 25-55 C	18 - 72 nrs at 30 - 35°C	48 - 96 hrs at 25°C							
Typical Colony Appearance	Clear to white colonies; some may produce pigment.	Colonies appear clear to creamy white; some may produce pigment. Tryptone Glucose Extract Broth with Indicator (TTC) produce red colonies.	Clear to white colonies; some may produce pigment.	Morphology and color vary depending on the microorganisms caught on the membrane. The majority of microorganisms develop pink to red colonies (formation from TTC indicator).	NOTE: All ampo Nutrient pads n	ules need to be stored at 2-8°C. eed to be stored at room temperature.					
pH at 25 °C	7.1 ±0.2	7.0 ± 0.2	7.3 ±0.2	7.1 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	For more ⁻ www.milli 1-800-MIL	For more technical information visit: www.millipore.com/dr_media 1-800-MILLIPORE (1-800-645-5476)		Se	Selection Guide		
Packaging Type	Non-luer tip ampoule	Non-luer tip ampoule	Luer tip ampoule	Dehydrated nutrient pad in 47mm dish							
Bacterial											
Selective	Wallerstein Differential Broth (WLD)	Pseudomonas Selective Broth	Lactose TTC Tergitol Media	MRS Media	m-Endo Total Coliform Broth	m-ColiBlue24 [®] Broth	Pseudomonas CN Media	m-FC Broth			
Catalogue Number	MHA000P2D (50/Pk) MX00WD220 (20/Pk)	MHA000P2P (50/Pk)	NPECC0150 (150/Pk) EZPDLT150 (150/Pk + 150 EZ-Pak [®] membranes	NPMRS0150 (150/Pk) MHAOOMRS2 (50/Pk)	MHAOOOP2E (50/Pk)	MOOPMCB24 (50/Pk)	NPPCN0150 (150/Pk)	MHA000P2F (with Rosolic Acid) MHA00FCR2 (without Rosolic Acid)			
Application	Used in breweries to detect and enumerate bacteria present in small numbers in a mixed flora sample. Used in conjunction with WLN broth. Cycloheximide inhibits the growth of most yeast and mold, allowing bacteria to grow.	Used for the detection of <i>Pseudomonas</i> species.	Used for the detection and enumeration of coliform organisms, thermotolerant coliform organisms and presumptive <i>Escherichia coli</i> in water, food and other samples.	Used for the isolation and enumeration of lactic acid bacteria species in food and other samples.	Used for the recovery of <i>E. coli</i> and coliform organisms in potable waters.	Used to detect both total coliforms and <i>E. coli</i> in water and beverages. This broth contains special inhibitors that prevent the growth of non-coliform bacteria but does not inhibit the growth of stressed organisms.	Used for the detection and enumeration of <i>Pseudomonas aeruginosa</i> in water.	Enumeration of fecal coliforms by membrane filtration technique at an elevated temperature for waste or effluent water.			
Incubation Time & Temperature	48 - 72 hrs at 30 - 35℃	24 – 72 hrs at 25 – 35°C	24 – 48 hrs at 35°C. For specific <i>E. coli</i> detection, 24 hrs at 44 ± 0.5°C.	3 – 5 days at 32.5°C in a 5% CO ₂ or in anaerobic atmosphere.	24 hrs at 35°C	24 hrs at 35°C	24 – 72 hrs at 35°C	24 hrs at 44.5°C			
Typical Colony Appearance	Bacteria appear small with green-blue color. If cycloheximide resistant yeast grow, they are creamy, green white.	All growth on this medium indicate the presence of <i>Pseudomonas</i> species. Colonies that are blue-green, brown or show fluorescence are presumptive <i>P. aeruginosa</i> .	Escherichia coli form small yellowish colonies (after 12–16 hours) which later change to orange and develop a yellow halo. Enterobacter and Klebsiella form yellow-green colonies. Salmonella, Proteus and Pseudomonas form red colonies with a bluish halo.	Colonies formed are small and white.	Red colonies with green metallic sheen.	<i>E. coli</i> form blue colonies and other coliforms form red colonies	Colonies formed are green-blue with blue halos and fluorescence under short wavelength (254 nm) ultraviolet light.	Fecal coliforms appear blue, other colonies appear gray to cream. In some rare cases, a membrane may have confirmed fecal coliforms that are pink in color.			
pH at 25 °C	5.5 ±0.2	7.1 +/-0.2	8.2 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	6.3 ± 0.2 NPMRS0150: (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	7.2 ±0.2	7.0 ±0.2	7.1 \pm 0.2 (with time, a slight reduction in may be noticed, which will not affect the recovery performance of the product)	7.4 ±0.2			
Packaging Type	MHA000P2D: Non-luer tip ampoule MX00WD220: Luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	NPMRS0150: Dehydrated nutrient pad in 47mm dish MHA00MRS2: Non-luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule			





Yeast and Mold	Orange Serum Media	Wallerstein Nutrient Broth (WI N)	m-Green/Schauffus Pottinger Media	Yeast & Mold Broth	Brettanomyces Broth	PRY (Preservative Resistant Yeast) Broth	Wort Media	Morris Orange Serum	Yeast and Mold Selective Broth	
Catalogue Number	MHA000P20 (50/Pk)	MHA00P2N (50/Pk)	MHA000P2M (50/Pk)	MX00YM220 (20/Pk)	MHA00BSM2 (50/Pk)	MHA00PRY2 (50/Pk)	NPWYM0150 (150/Pk)	MHA00MS02	MHA00P2SM (with Chloramphenicol) (50/Pk)	
Application	NPOSA0150 (150/Pk) Used for the isolation and enumeration of acidophilic and aciduric microorganisms in water, beverages and foods.	MXOOWN220 (20/Pk) Used to detect yeast and mold in worts, beers and other fermentation products. Used in cojunction with WLD.	A low pH culture medium used to detect yeast and mold and other aciduric organisms	Used for the isolation and cultivation of yeast, mold and other aciduric organisms.	Detection of <i>Brettanomyces</i> in wine and beer. Bacteria and other yeasts are inhibited.	A low pH selective culture medium for the detection of spoilage microorganisms resistant to acetic acid.	Used for the isolation and enumeration of yeast in beverages, beer and wine.	Used to detect yeast and mold resistant to sodium benzoate (like <i>Zygosaccharomyces bailii</i>) in beverage.	Used to detect yeast and mold. Chloramphenicol is used to suppress background bacterial growth.	
Incubation Time & Temperature	48 hrs – 5 days at 24 – 32°C	48 hrs – 5 days for yeast at 20°C 35°C for bacteria	48 – 72 hrs at 28 – 32°C	48 – 72 hrs at 20 – 30°C	5 – 7 days at 25°C	48 – 72 hrs at 30°C	48 hrs - 5 days at 22.5°C	2 -7 days at 25-30°C	48 - 72 hrs at 28 - 35 °C	
Typical Colony Appearance	Yeast appear white, creamy and large. Bacteria are smaller, white or transparent.	Mold can appear non-pigmented to white, with various texture. Yeast appear as creamy, white larger colonies. Bacteria appear blue-green.	Yeast are large green opaque colonies. Mold appears green and filamentous. Bacteria able to grow at this pH form smaller clear to white colonies.	Yeast produce white colonies with a creamy texture. Mold is rough textured and/or filamentous. Bacteria are smaller and clear to white.	Colonies appear small, white and creamy.	Yellow	Yeasts develop smooth white or colored colonies.	Yeast colonies appear as white, creamy and large colonies.	Yeast appear as large green and opaque colonion. Mold is green and filamentous.	
pH at 25 °C	MHA000P20: 5.6 \pm 0.2 NPOSA0150: 4.5 \pm 0.2 (with time, a slight reduction of pH may be noticed, which will not affect the recovery performance of the product)	5.5 ±0.2	MHA000P2M: 4.6 ±0.2 NPSPY0150: 4.5 ±0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	4.6 ±0.3	3.5 ±0.2	3.6 ±0.2	4.5 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recover performance of the product)	3.5 ± 0.2	4.4 ±0.2	
Packaging Type	MHA000P20: Non-luer tip ampoule NPOSA0150: Dehydrated nutrient pad in 47mm dish	MHAOOP2N: (Non-luer tip ampoule) MXOOWN220: (Luer tip ampoule)	MHA000P2M: (Non-luer tip ampoule) NPSPY0150: (Dehydrated nutrient pad in 47mm dish)	Luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule	Non-luer tip ampoule	
Total Viable Organism/Total Viable Count	<image/>	Tryptone Glucose Extract (TGE) Broth	Tryptic Soybean Broth (TSB)	Standard Count Media						
Catalogue Number	MHA000P2S (50/Pk)	MHA000P2T (50/Pk) With TTC Indicator: MHA00P2TT (50/Pk)	MX00TT220 (20/Pk)	NPSTC0150 (150/Pk)			ТМ	Σ		
Application	Recovery of heterotrophic bacteria found in various types of water, especially high-purity or potable. Prior	A non-selective medium to detect total heterotrophic microorganisms in water	Used to detect total heterotrophic microorganisms in water and other liquids.	Used for the cultivation of fastidious and other microorganisms found in water, wastewater,						
Incubation Time	to use, warm the media at 30-50 °C until liquefied. 48 – 72 hrs at 25 – 35°C	and other liquids. 48 - 72hrs at 25-35°C	18 - 72 hrs at 30 - 35°C	raw materials, beverages, beer, food, etc. 24 - 48 hrs at 35°C						
& Temperature Typical Colony Appearance	Clear to white colonies; some may produce pigment.	Colonies appear clear to creamy white; some may produce pigment. Tryptone Glucose Extract Broth with Indicator (TTC) produce red colonies.	Clear to white colonies; some may produce pigment.	48 - 96 hrs at 25°C Morphology and color vary depending on the microorganisms caught on the membrane. The majority of microorganisms develop pink to red colonies (formation from TTC indicator)	NOTE: All ampo Nutrient pads r	pules need to be stored at 2-8°C. need to be stored at room temperature.				
pH at 25 °C	7.1 ±0.2	7.0 ± 0.2	7.3 ±0.2	7.1 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	For more technical information visit: www.millipore.com/dr_media 1-800-MILLIPORE (1-800-645-5476)		wth Media Selection Guid			
Packaging Type	Non-luer tip ampoule	Non-luer tip ampoule	Luer tip ampoule	Dehydrated nutrient pad in 47mm dish						
Bacterial										
Selective	Wallerstein Differential Broth (WLD)	Pseudomonas Selective Broth	Lactose TTC Tergitol Media	MRS Media	m-Endo Total Coliform Broth	m-ColiBlue24® Broth	Pseudomonas CN Media	m-FC Broth		
Catalogue Number	MHA000P2D (50/Pk) MX00WD220 (20/Pk)	MHA000P2P (50/Pk)	NPECC0150 (150/Pk) EZPDLT150 (150/Pk + 150 EZ-Pak [®] membranes	NPMRS0150 (150/Pk) MHA00MRS2 (50/Pk)	MHA000P2E (50/Pk)	MOOPMCB24 (50/Pk)	NPPCN0150 (150/Pk)	MHA000P2F (with Rosolic Acid) MHA00FCR2 (without Rosolic Acid)		
Application	Used in breweries to detect and enumerate bacteria present in small numbers in a mixed flora sample. Used in conjunction with WLN broth. Cycloheximide inhibits the growth of most yeast and mold, allowing bacteria to grow.	Used for the detection of <i>Pseudomonas</i> species.	Used for the detection and enumeration of coliform organisms, thermotolerant coliform organisms and presumptive <i>Escherichia coli</i> in water, food and other samples.	Used for the isolation and enumeration of lactic acid bacteria species in food and other samples.	Used for the recovery of <i>E. coli</i> and coliform organisms in potable waters.	Used to detect both total coliforms and <i>E. coli</i> in water and beverages. This broth contains special inhibitors that prevent the growth of non-coliform bacteria but does not inhibit the growth of stressed organisms.	Used for the detection and enumeration of <i>Pseudomonas aeruginosa</i> in water.	Enumeration of fecal coliforms by membrane filtration technique at an elevated temperature for waste or effluent water.		
Incubation Time & Temperature	48 – 72 hrs at 30 – 35°C	24 - 72 hrs at 25 - 35℃	24 – 48 hrs at 35°C. For specific <i>E. coli</i> detection, 24 hrs at 44 ± 0.5°C.	3 – 5 days at 32.5°C in a 5% CO ₂ or in anaerobic atmosphere.	24 hrs at 35°C	24 hrs at 35°C	24 - 72 hrs at 35°C	24 hrs at 44.5°C		
Typical Colony Appearance	Bacteria appear small with green-blue color. If cycloheximide resistant yeast grow, they are creamy, green white.	All growth on this medium indicate the presence of <i>Pseudomonas</i> species. Colonies that are blue-green, brown or show fluorescence are presumptive <i>P. aeruginosa</i> .	Escherichia coli form small yellowish colonies (after 12–16 hours) which later change to orange and develop a yellow halo. Enterobacter and Klebsiella form yellow-green colonies. Salmonella, Proteus and Pseudomonas form red colonies with a bluish halo.	Colonies formed are small and white.	Red colonies with green metallic sheen.	<i>E. coli</i> form blue colonies and other coliforms form red colonies	Colonies formed are green-blue with blue halos and fluorescence under short wavelength (254 nm) ultraviolet light.	Fecal coliforms appear blue, other colonies appear gray to cream. In some rare cases, a membrane may have confirmed fecal coliforms that are pink in color.		
pH at 25 °C	5.5 ±0.2	7.1 +/-0.2	8.2 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	6.3 ± 0.2 NPMRS0150: (with time, a slight reduc- tion in pH may be noticed, which will not affect the recovery performance of the product)	7.2 ±0.2	7.0 ±0.2	7.1 ±0.2 (with time, a slight reduction in may be noticed, which will not affect the recovery performance of the product)	7.4 ±0.2		
Packaging Type	MHA000P2D: Non-luer tip ampoule MX00WD220: Luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	NPMRS0150: Dehydrated nutrient pad in 47mm dish MHA00MRS2: Non-luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule		





Yeast and Mold	Orange Serum Media	Wallerstein Nutrient Broth (WLN)	m-Green/Schauffus Pottinger Media	Yeast & Mold Broth	Brettanomyces Broth	PRY (Preservative Resistant Yeast) Broth	Wort Media	Morris Orange Serum	Yeast and Mold Selective Broth	
Catalogue Number	MHA000P20 (50/Pk) NP0SA0150 (150/Pk)	MHA00P2N (50/Pk) MX00WN220 (20/Pk)	MHA000P2M (50/Pk) NPSPY0150 (150/Pk)	MX00YM220 (20/Pk)	MHAOOBSM2 (50/Pk)	MHAOOPRY2 (50/Pk)	NPWYM0150 (150/Pk)	MHA00MS02	MHA00P2SM (with Chloramphenicol) (50/Pk)	
Application	Used for the isolation and enumeration of acidophilic and aciduric microorganisms in water, beverages and foods.	Used to detect yeast and mold in worts, beers and other fermentation products. Used in cojunction with WLD.	A low pH culture medium used to detect yeast and mold and other aciduric organisms	Used for the isolation and cultivation of yeast, mold and other aciduric organisms.	Detection of <i>Brettanomyces</i> in wine and beer. Bacteria and other yeasts are inhibited.	A low pH selective culture medium for the detection of spoilage microorganisms resistant to acetic acid.	Used for the isolation and enumeration of yeast in beverages, beer and wine.	Used to detect yeast and mold resistant to sodium benzoate (like <i>Zygosaccharomyces bailii</i>) in beverage.	Used to detect yeast and mold. Chloramphenicol is used to suppress background bacterial growth.	
Incubation Time & Temperature	48 hrs – 5 days at 24 – 32°C	48 hrs – 5 days for yeast at 20°C 35°C for bacteria	48 – 72 hrs at 28 – 32°C	48 – 72 hrs at 20 – 30°C	5 – 7 days at 25°C	48 – 72 hrs at 30°C	48 hrs − 5 days at 22.5°C	2 -7 days at 25-30°C	48 – 72 hrs at 28 – 35 °C	
Typical Colony Appearance	Yeast appear white, creamy and large. Bacteria are smaller, white or transparent.	Mold can appear non-pigmented to white, with various texture. Yeast appear as creamy, white larger colonies. Bacteria appear blue-green.	Yeast are large green opaque colonies. Mold appears green and filamentous. Bacteria able to grow at this pH form smaller clear to white colonies.	Yeast produce white colonies with a creamy texture. Mold is rough textured and/or filamentous. Bacteria are smaller and clear to white.	Colonies appear small, white and creamy.	Yellow	Yeasts develop smooth white or colored colonies.	Yeast colonies appear as white, creamy and large colonies.	Yeast appear as large green and opaque colonio Mold is green and filamentous.	
pH at 25 °C	MHA000P20: 5.6 \pm 0.2 NP0SA0150: 4.5 \pm 0.2 (with time, a slight reduction of pH may be noticed, which will not affect the recovery performance of the product)	5.5 ±0.2	MHA000P2M: 4.6 ±0.2 NPSPY0150: 4.5 ±0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	4.6 ±0.3	3.5 ±0.2	3.6 ±0.2	4.5 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	3.5 ± 0.2	4.4 ±0.2	
Packaging Type	MHA000P20: Non-luer tip ampoule	MHA00P2N: (Non-luer tip ampoule)	MHA000P2M: (Non-luer tip ampoule)	Luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule	Non-luer tip ampoule	
Total Viable Organism/Total Viable Count	Heterotrophic Plate Count (HPC) Broth	Tryptone Glucose Extract (TGE) Broth	Tryptic Soybean Broth (TSB)	Standard Count Media						
Catalogue Number	MHA000P2S (50/Pk)	MHA000P2T (50/Pk) With TTC Indicator: MHA00P2TT (50/Pk)	MX00TT220 (20/Pk)	NPSTC0150 (150/Pk) With TTC Indicator: NPTTC0150 (150/Pk)			ТМ	Σ		
Application	Recovery of heterotrophic bacteria found in various	A non-selective medium to detect total	Used to detect total heterotrophic	Used for the cultivation of fastidious and other						
	types of water, especially high-purity or potable. Prior to use, warm the media at 30-50 °C until liquefied.	heterotrophic microorganisms in water and other liquids.	microorganisms in water and other liquids.	microorganisms found in water, wastewater, raw materials, beverages, beer, food, etc.						
Incubation Time & Temperature	48 – 72 hrs at 25 – 35°C	48 - 72hrs at 25-35°C	18 – 72 hrs at 30 – 35°C	24 – 48 hrs at 35°C 48 – 96 hrs at 25°C						
Typical Colony Appearance	Clear to white colonies; some may produce pigment.	Colonies appear clear to creamy white; some may produce pigment. Tryptone Glucose Extract Broth with Indicator (TTC) produce red colonies.	Clear to white colonies; some may produce pigment.	Morphology and color vary depending on the microorganisms caught on the membrane. The majority of microorganisms develop pink to red colonies (formation from TTC indicator).	NOTE: All ampoules need to be stored at 2-8°C. Nutrient pads need to be stored at room temperature.					
pH at 25 °C	7.1 ±0.2	7.0 ± 0.2	7.3 ±0.2	7.1 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	For more www.milli 1-800-MIL	For more technical information visit: www.millipore.com/dr_media 1-800-MILLIPORE (1-800-645-5476)		MICRODIAL GROWTH MEDIA Selection Gui		
Packaging Type	Non-luer tip ampoule	Non-luer tip ampoule	Luer tip ampoule	Dehydrated nutrient pad in 47mm dish						
Bacterial										
Selective	Wallerstein Differential Broth (WLD)	Pseudomonas Selective Broth	Lactose TTC Tergitol Media	MRS Media	m-Endo Total Coliform Broth	m-ColiBlue24® Broth	Pseudomonas CN Media	m-FC Broth		
Catalogue Number	MHA000P2D (50/Pk) MX00WD220 (20/Pk)	MHA000P2P (50/Pk)	NPECC0150 (150/Pk) EZPDLT150 (150/Pk + 150 EZ-Pak®membranes	NPMRS0150 (150/Pk) MHA00MRS2 (50/Pk)	MHA000P2E (50/Pk)	MOOPMCB24 (50/Pk)	NPPCN0150 (150/Pk)	MHA000P2F (with Rosolic Acid) MHA00FCR2 (without Rosolic Acid)		
Application	Used in breweries to detect and enumerate bacteria present in small numbers in a mixed flora sample. Used in conjunction with WLN broth. Cycloheximide inhibits the growth of most yeast and mold, allowing bacteria to grow.	Used for the detection of <i>Pseudomonas</i> species.	Used for the detection and enumeration of coliform organisms, thermotolerant coliform organisms and presumptive <i>Escherichia coli</i> in water, food and other samples.	Used for the isolation and enumeration of lactic acid bacteria species in food and other samples.	Used for the recovery of <i>E. coli</i> and coliform organisms in potable waters.	Used to detect both total coliforms and <i>E. coli</i> in water and beverages. This broth contains special inhibitors that prevent the growth of non-coliform bacteria but does not inhibit the growth of stressed organisms.	Used for the detection and enumeration of <i>Pseudomonas aeruginosa</i> in water.	Enumeration of fecal coliforms by membrane filtration technique at an elevated temperature for waste or effluent water.		
Incubation Time & Temperature	48 – 72 hrs at 30 – 35°C	24 - 72 hrs at 25 - 35°C	24 – 48 hrs at 35°C. For specific <i>E. coli</i> detection, 24 hrs at 44 ± 0.5°C.	3 – 5 days at 32.5°C in a 5% CO ₂ or in anaerobic atmosphere.	24 hrs at 35°C	24 hrs at 35°C	24 – 72 hrs at 35°C	24 hrs at 44.5°C		
Typical Colony Appearance	Bacteria appear small with green-blue color. If cycloheximide resistant yeast grow, they are creamy, green white.	All growth on this medium indicate the presence of <i>Pseudomonas</i> species. Colonies that are blue-green, brown or show fluorescence are presumptive <i>P. aeruginosa</i> .	Escherichia coli form small yellowish colonies (after 12–16 hours) which later change to orange and develop a yellow halo. Enterobacter and Klebsiella form yellow-green colonies. Salmonella, Proteus and Pseudomonas form red colonies with a bluish halo.	Colonies formed are small and white.	Red colonies with green metallic sheen.	<i>E. coli</i> form blue colonies and other coliforms form red colonies	Colonies formed are green-blue with blue halos and fluorescence under short wavelength (254 nm) ultraviolet light.	Fecal coliforms appear blue, other colonies appear gray to cream. In some rare cases, a membrane may have confirmed fecal coliforms that are pink in color.		
pH at 25 °C	5.5 ±0.2	7.1 +/-0.2	8.2 ± 0.2 (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	6.3 ± 0.2 NPMRS0150: (with time, a slight reduction in pH may be noticed, which will not affect the recovery performance of the product)	7.2 ±0.2	7.0 ±0.2	7.1 ±0.2 (with time, a slight reduction in may be noticed, which will not affect the recovery performance of the product)	7.4 ±0.2		
Packaging Type	MHA000P2D: Non-luer tip ampoule MX00WD220: Luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	NPMRS0150: Dehydrated nutrient pad in 47mm dish MHA00MRS2: Non-luer tip ampoule	Non-luer tip ampoule	Non-luer tip ampoule	Dehydrated nutrient pad in 47mm dish	Non-luer tip ampoule		







